

Communications to the editor

A NOVEL β -LACTAMASE
INHIBITOR, SF-2103 A
PRODUCED BY A *STREPTOMYCES**

Sir:

A novel β -lactamase inhibitor, SF-2103 A, was discovered in culture filtrates of a new streptomycete strain SF-2103 for which was proposed the name *Streptomyces sulfonofaciens* sp. nov. In this communication, we report the isolation and structural elucidation of SF-2103 A.

The inhibitor was produced by submerged cultivation of strain SF-2103 in a 2,000-liter fermentor containing 1,000 liters of a medium consisting of 2% maltose syrup, 1.2% soybean meal, 1.2% wheat germ, 0.05% Na_2SO_4 , 0.001% $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$ and 0.1% CaCO_3 . Fermentation was carried out under agitation at 28°C for 86 hours, and the maximum titer was 12 μg of SF-2103 A per ml, based on inhibitory activity toward the β -lactamase of *Proteus vulgaris* GN-75¹⁾.

The inhibitor in the culture filtrate (900 liters) was extracted with dichloromethane (300 liters) containing 0.2% benzyldimethyltetradecylammonium chloride at pH 6.1 and retransferred into 1% NaI aqueous solution (28 liters). The aqueous extract was passed through a column of active charcoal (1 liter, Wako Pure Chemical Industries), and the effluent was charged on a column of DEAE-Sephadex A-25 (3 liters). After washing with 0.02 M phosphate buffer containing 0.2 M

NaCl (30 liters, pH 7.2), SF-2103 A was eluted with the same buffer containing 0.3 M NaCl. The active eluate was desalted by adsorption on a column of active charcoal (4 liters) and eluted with 50% aqueous acetone. The eluate was subjected to column chromatography with DEAE-Sephadex A-25 (500 ml) developed with 0.02 M phosphate buffer containing 0.2 M NaCl. The active eluate was desalted in the same manner as above, and applied to a column of Sephadex G-10 (800 ml) developed with water. Evaporation and lyophilization of the active effluent yielded the sodium salt of pure SF-2103 A (510 mg).

SF-2103 A sodium salt is a colorless powder which decomposes gradually over 168°C, and is soluble in water, dimethyl sulfoxide and *N,N*-dimethylformamide but insoluble in lower alcohols, acetone, ethyl acetate and benzene. It shows positive color reactions with GREIG-LEABACK's and EHRlich's reagents, and negative with ninhydrin. SF-2103 A showed UV maxima (0.02 M phosphate buffer at pH 7.2) as follows: λ_{max} 230 nm (ϵ 5416), 266 nm (ϵ 5783) (extinguishable by addition of NH_2OH); IR (KBr) 1755 (β -lactam), 1610 (amide), 1220 cm^{-1} (S=O); $[\alpha]_{\text{D}}^{20}$ -16.3° (*c* 1, water); CD $[\theta]_{203}$ 0, $[\theta]_{232}$ $-132,800$, $[\theta]_{248}$ 0, $[\theta]_{264}$ $+51,400$, $[\theta]_{310}$ 0 (*c* 0.01, water).

Anal. Calcd. for $\text{C}_9\text{H}_9\text{NO}_{10}\text{S}_2\text{Na}_3 \cdot 2\text{H}_2\text{O}$:

C 23.53, H 2.61, N 3.05, S 13.94, Na 15.03.

Found: C 23.62, H 2.44, N 3.00, S 13.81, Na 14.97.

Table 1. ^1H NMR chemical shifts of SF-2103 A (**1**) and non β -lactam compounds **2a** and **2b**.

Proton	δ ppm (<i>J</i> Hz)		
	1	2a	2b
9-CH ₃	1.60 (d, 3H, $J_{8,9}=6.4$)	1.52 (d, 3H, $J=6$)	1.53 (d, 3H, $J=6$)
8-CH	5.01 (m, 1H, $J_{8,9}=8.8$)	4.76 (m, 1H)	4.93 (m, 1H, $J=6$)
6-CH	3.99 (dd, 1H, $J_{6,5}=5.9$)	2.92 (t, 1H, $J=8$)	2.98 (dd, 1H, $J=6$)
5-CH	4.54 (m, 1H, $J_{5,4}=10.3$, $J_{5,4'}=9.0$)	4.61 (m, 1H)	4.33 (m, 1H)
4-CH ₂	3.06 (dd, 1H, $J_{4,4'}=16.9$) 3.48 (dd, 1H)	2.54 (t, 2H, $J=7$)	2.05 (dd, 1H, $J=7$) 2.66 (dd, 1H, $J=8$)
3-CH		4.69 (m, 1H)	~ 4.6 (1H)

200 MHz, D₂O, DSS as external reference.

* This work was presented in the 225th Scientific Meeting of Japan Antibiotics Research Association, Tokyo, Sept. 25, 1981

Table 2. ^{13}C NMR chemical shifts of SF-2103 A (1) and non β -lactam compound (2).

Assignment	δ ppm			
	1		2	
9 $-\text{CH}_3$	19.5	q	18.9,	17.7 q
4 $-\text{CH}_2-$	33.0	t	30.7	t
6 $>\text{CH}-$	55.2	d	55.9,	60.2 d
5 $>\text{CH}-$	58.9	d	72.2,	71.5 d
8 $>\text{CH}-$	73.8	d	77.6,	77.4 d
3 $>\text{C}=(\text{CH}-)$	129.2*	s	69.9,	69.7 d
2 $>\text{C}=\text{O}$	139.4*	s	171.6*	s
7 $>\text{C}=\text{O}$	168.9**	s	170.9*,	170.5* s
10 $>\text{C}=\text{O}$	178.2**	s	178.7*,	178.8* s

50 MHz, D_2O , DSS as external reference.

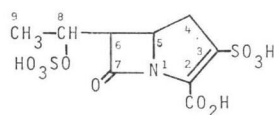
*, **: Assignments may be interconverted within each column.

The UV and IR absorption of SF-2103 A suggested the presence of a 1-carba-2-penem-3-carboxylate structure²⁾. ^1H NMR and ^{13}C NMR spectra of SF-2103 A (Tables 1 and 2) showed close resemblance to those of MM 4550³⁾ and MC 696-SY2-A⁽⁴⁾ except for their C-3 side chain. On high-voltage paper electrophoresis at 2,800V for 15 minutes in pyridine-acetate buffer (pH 6.4), SF-2103 A migrated toward the anode at the rate of R_m (relative mobility to MC696-SY2-A*) 1.6. Thus, the polyacidic nature of SF-2103 A coupled with elemental analysis suggested the presence of a sulfur-containing function, such as sulfate or sulfonate instead of the neutral C-3 side chain of MC696-SY2-A. Hydrolysis of SF-2103 A upon refluxing in 0.5 N HCl for 20 hours gave one mole of sulfate as analysed by gravimetric analysis.

From the physico-chemical properties and spectral data mentioned above, the structure of SF-2103 A was proposed to be 1 as shown in Scheme 1. The orientation of H-5 and H-6 in SF-2103 A was deduced to be *cis* from the vicinal coupling constants, $J_{5,6} = 5.9$ Hz.

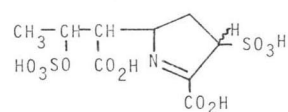
The presence of the sulfonate rather than the sulfate group at C-3 was confirmed by the following degradation. Mild hydrolysis in 0.02 M acetate buffer of pH 4.0 at 36°C for 20 hours yielded a non β -lactam compound (2), colorless powder, mp: gradually decomposed over 182°C, $[\alpha]_D^{20} -54^\circ$ (*c* 1, water), no characteristic UV absorption, no IR band around 1755 cm^{-1} .

* MC696-SY2-A was kindly supplied by Dr. K. MAEDA, Institute of Microbial Chemistry, Japan.



SF-2103 A (1)

pH 4.0
(0.02M acetate buffer)
36°C, 20 hours



2

Anal. Calcd. for $\text{C}_9\text{H}_9\text{NO}_{11}\text{S}_2\text{Na}_4 \cdot 2\text{H}_2\text{O}$:

C 21.64, H 2.61, N 2.81, S 12.82.

Found: C 21.93, H 2.94, N 2.82, S 12.87.

Most signals of the ^1H NMR and ^{13}C NMR spectra of 2 (Tables 1 and 2) were observed as pairs, suggesting that 2 was a mixture of diastereomers 2a and 2b (Table 1)⁴⁾. Based on the ^1H NMR spectrum of 2, the ratio of 2a : 2b was approximately 2 : 1. A pair of signals of C-3 (δ 69.7, 69.9 ppm) was observed at a higher field ($\Delta\delta$ 7~8 ppm) than that of C-8 (δ 77.4, 77.6 ppm) bearing a sulfate ester group. This result suggests that C-3 links to a sulfonate group.

SF-2103 A possessing a sulfonate group was obviously distinguishable from known carba-

Table 3. Antibacterial spectrum of SF-2103 A.

Organism	MIC ($\mu\text{g/ml}$)
<i>Staphylococcus aureus</i> 209P JC-1	25
<i>Staphylococcus aureus</i> Smith	25
<i>Streptococcus faecalis</i> ATCC 8043	50
<i>Bacillus subtilis</i> ATCC 6633	50
<i>Escherichia coli</i> NIHJ JC-2	12.5
<i>Escherichia coli</i> No. 29	6.25
<i>Klebsiella pneumoniae</i> PCI 602	50
<i>Salmonella typhi</i> O-901-W	12.5
<i>Salmonella enteritidis</i> No. 11	50
<i>Shigella dysenteriae</i> Shigae	3.13
<i>Proteus vulgaris</i> OX-19	25
<i>Proteus morgani</i> Kōno	50
<i>Serratia marcescens</i> No. 2	100
<i>Pseudomonas aeruginosa</i> E-2	>100
<i>Pseudomonas cepacia</i> M-0527	100

Table 4. β -Lactamase-inhibitory activities of SF-2103 A.

Source of β -lactamase ¹⁾	Substrate	I_{50} (μ g/ml)*
<i>Staphylococcus aureus</i> MS258	PCG**	> 50 (4.0)
<i>Escherichia coli</i> W3630 RGN823	PCG	1.6 (0.05)
<i>Proteus vulgaris</i> GN76/C-1	CET**	0.0036 (0.00015)
<i>Citrobacter freundii</i> GN346	CET	0.0086 (0.00068)

* Concentrations required to cause 50% inhibition were determined by the modified microiodometric method.⁸⁾ The values in parentheses were obtained when inhibitor was preincubated with β -lactamase for 10 minutes at 30°C before substrate addition.

** PCG and CET are penicillin G and cephalothin, respectively.

penem antibiotics such as the olivanic acids⁹⁾, epithienamycins⁹⁾ and carpetimycins⁷⁾.

SF-2103 A exhibits a broad antibacterial spectrum with moderate activity against Gram-positive and Gram-negative bacteria (Table 3). However, it shows powerful inhibition against both types of enzymes of penicillinases and cephalosporinases as shown in Table 4.

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